DEPARTMENT OF CHEMISTRY

UNIVERSITY OF OXFORD

The Lego® Spectrophotometer

Practical Investigations of Dye

Investigating Quinoline Yellow WS Dye (E104)

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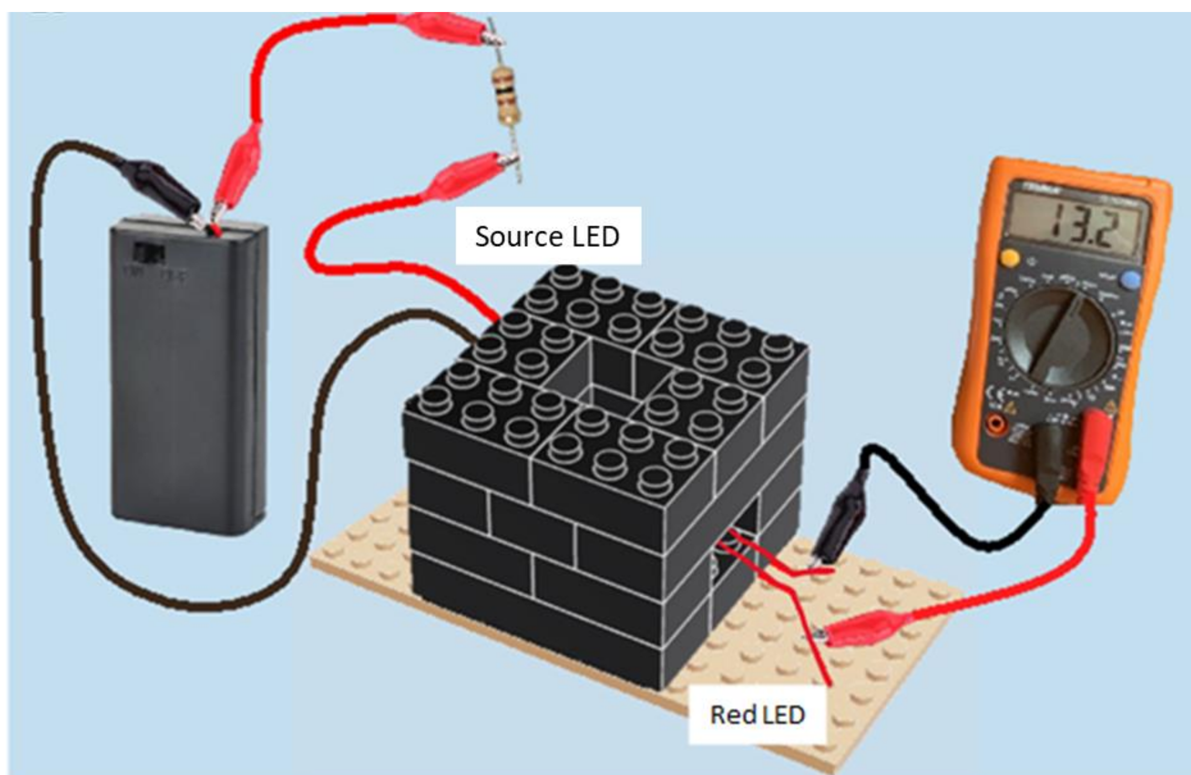
Introduction:

The Lego® Spectrophotometer is a simple Lego® apparatus which can be used for a single beam scanning spectrophotometer experiment. It can measure the concentration of unknown substances, determine the proportions of substances in a mixture (given the component absorbance spectra do not overlap), and monitor the kinetics of a reaction.



Spectrophotometry

Spectrophotometry is a type of electromagnetic spectroscopy which allows determination of the absorptivity or transmittivity of a substance under investigation. Spectrophotometry is commonly used in:

1. Drug testing – e.g., to determine if an athlete has misused anabolic steroids
2. Coffee brewing – e.g., to determine if coffee meets industry standard to ensure consistent coffee flavour and strength.
3. Plastics manufacturing – e.g., to ensure even colour distribution (likely used by Lego® to ensure uniform brick colours!).
4. Paint colour mixing – e.g., determining correct opacity of paint.

Glossary: Absorptivity is a measure of how strongly a species absorbs light at a specific wavelength. This is denoted 'ε' in the Beer law (see later).
Transmittivity is a measure of how much light of a specific wavelength a species lets through.

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Many applications of spectrophotometry rely on the colour of a substance, but why is this? First, we need to understand how spectrophotometers work.

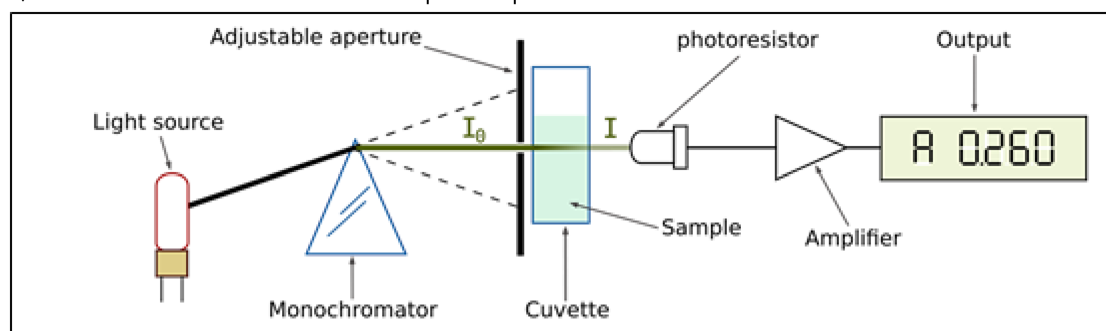

Figure 1: Schematic layout of a single beam scanning spectrophotometer.

Spectrophotometer Design

All spectrophotometers have a similar design illustrated above (Figure 1). Their basic mechanism is outlined below:

1. A light source emits light with a range of wavelengths. This type of light is polychromatic ('many-coloured').
2. The polychromatic light source passes through a monochromator, which refocuses only one wavelength onto the cuvette.
3. The focussed light beam passes through the sample with incident intensity, I_0. The sample absorbs some light and transmits the rest, with transmission intensity, I. Since some light is absorbed by the sample, $I_0 > I$.
4. The light with transmitted intensity, I, reaches the detector which displays an output of voltage.
5. The resultant voltage can be used to deduce the transmitted intensity. Voltage, V, is directly proportional to transmitted intensity, I.

Coloured solutions absorb light in the visible region, so spectrophotometry is an excellent technique for analysing such samples. In these cases, the light source can be a coloured bulb.

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Why are Objects Coloured?

The colour of a compound or solution arises from its interaction with visible light. Visible light is the portion of the electromagnetic spectrum we can see, with wavelengths between 400 and 700 nm (Figure 2).

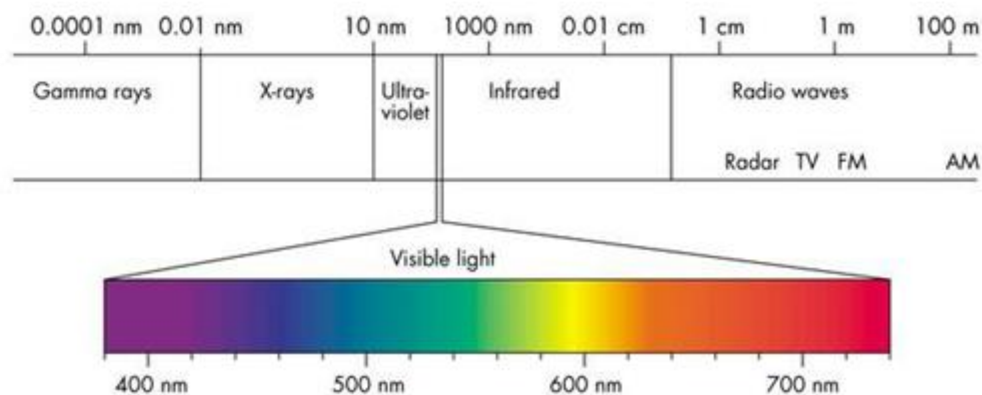

Figure 2: A summary of the electromagnetic spectrum. The wavelengths of the different coloured components of visible light are indicated.

Most objects will absorb some wavelengths of light better than others and the colour of a substance depends on the wavelengths of light that it absorbs. For a solution to appear as a particular colour, the dissolved compound must absorb the complementary colour to which it appears. Complementary colours lie opposite on a colour wheel (Figure 3). For example, red is the complementary colour of green, and blue and orange are complementary colours.

Glossary: Complementary colours are colours that are opposite on the colour wheel, below. For example, red is the complementary colour of green, and blue and orange are complementary colours.

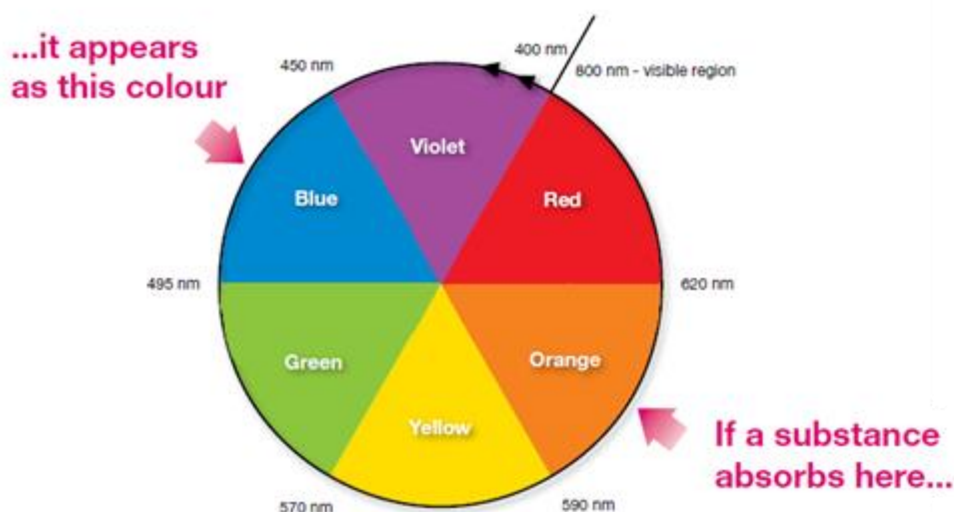

Figure 3: A colour wheel illustrating the concept of complementary colours.

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The interaction of light with the sample is an important consideration in the Lego® spectrophotometer. We must choose a light source that the compound will absorb in order to record accurate optical measurements. To investigate different coloured dyes you need to choose a correctly coloured LED light source. In the case of quinoline yellow, which appears yellow, the complementary colour is blue and a blue LED is employed as the light source.

Why Do Dye Molecules Absorb Light?

Coloured compounds often contain conjugated systems.

Glossary: A **conjugated** system is one where the bonds alternate between single and double bonds, for example between carbon atoms.

Electrons in compounds can be excited from the HOMO (highest occupied molecular orbital) to the LUMO (lowest unoccupied molecular orbital) by absorbing energy in the form of a photon (see Figure 4). In the case of many conjugated systems, the photon (packet of energy) absorbed to excite an electron lies in the visible light region, and so the compound appears coloured. The wavelength of the absorbed photon will correspond to the complementary colour of the compound

Glossary: The **HOMO** is the Highest Occupied Molecular Orbital. It is the highest energy level an electron can be in, in a molecule that is in its lowest energy state (ground state). The **LUMO** is the Lowest Unoccupied Molecular Orbital, which is the molecular orbital of next highest energy. This is empty when the molecule is in its ground state (state of lowest possible energy).

Beer-Lambert Law

As described above, a fraction of light passing through a sample is absorbed so that:

$$I_t = I_0 - I_{abs}$$

Where I_t = intensity of light *transmitted* by the sample, I_0 = intensity of incident light *hitting* the sample and I_{abs} = intensity of light *absorbed* by the sample.

The **Lambert Law** states that:

$$I_t = I_0 - I_{abs} = I_0 \times 10^{-A}$$

Where A = sample absorbance.

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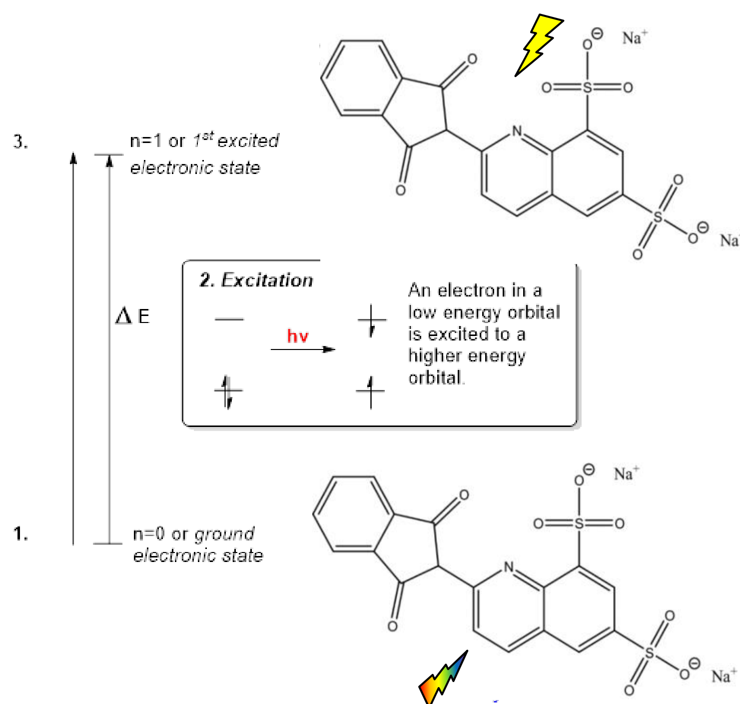

Figure 4: Excitation of an electron in Quinoline Yellow by visible light. 1. White visible light, which contains all colours (wavelengths) of light, is incident on the dye molecule. 2. The photons with a wavelength that corresponds to the right amount of energy (the difference between the ground state and excited state or ΔE) is absorbed. 3. The photons with wavelengths that are not strongly absorbed are reflected. To an observer, the dye appears to be the complementary colour to the strongly absorbed colour, so it appears yellow.

We can rearrange the Lambert Law equation:

$$\frac{I_t}{I_0} = 10^{-A}$$

And take logarithms of both sides:

$$A = -\log_{10}\left(\frac{I_t}{I_0}\right)$$

This equation will be used later on to calculate the absorbance (A) of dye solutions.

The **Beer Law** states that:

$$A = \varepsilon c l$$

Where c = concentration (normally in mol dm^{-3}), l = path length (1 cm for standard cuvettes) and ε = absorption coefficient (normally in mol^{-1} dm^{3} cm^{-1}).

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Therefore, a linear plot of A (sample absorbance) against c (sample concentration), yields a straight line with a gradient equal to εl. You will use this later on to determine the concentration of dye in an unknown sample

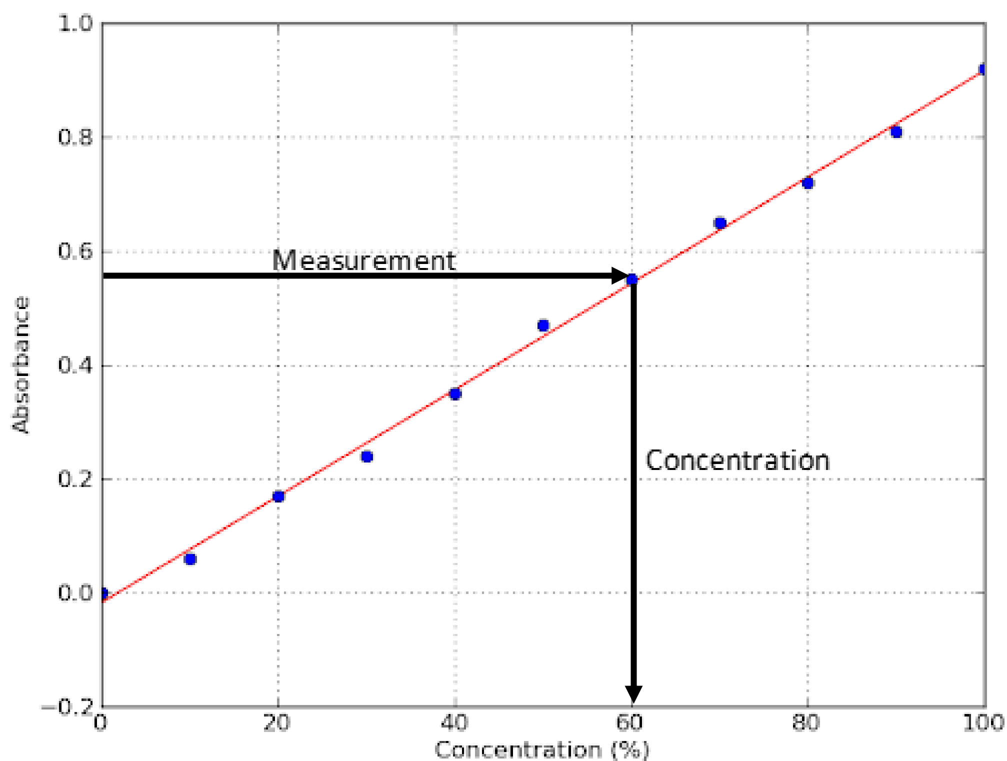

Figure 5: Concentration calibration plot to determine unknown concentration.

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An Investigation of Quinoline Yellow WS Dye:

Objectives:

- To gain an understanding of how spectrophotometers work.
- To apply simple circuitry to build a functional spectrophotometer.
- To plot a calibration curve for Quinoline Yellow WS dye
- To determine the concentration of Quinoline Yellow in an unknown solution.

Background: Quinoline Yellow Dye Background

Quinoline Yellow Dye WS (E number E104) is a water-soluble dye. This is a result of the sulfonate groups found on the terminal hexagonal ring. Quinoline Yellow SS does not have these groups and is therefore insoluble in water.

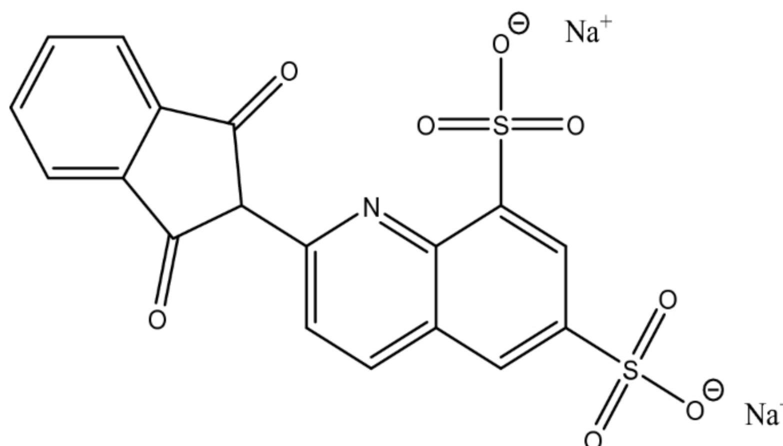

Figure 6: The Structure of Quinoline Yellow SS

This dye consists of an extended conjugate system. However, as this dye is a bright yellow colour, it instead absorbs light in the blue region of 450-495 nm. Thus, we know a HOMO to LUMO electron transition must occur at an energy corresponding to a wavelength in this region.

Quinoline Yellow WS is commonly used as a food additive in the United Kingdom and Europe, with a recommended maximum daily usage of 0.5 mg/kg of body weight. In the United States it is not approved by the FDA (Food and Drug Administration) for use as a food additive but is found in many medicines and cosmetics.

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Determination of an Unknown Concentration:

In this part of the practical, you will determine the concentration of Quinoline Yellow WS dye in a sample of cough sweets of unknown concentration.

From the introduction, remember that:

$$A(c) = -\log_{10}\left(\frac{I_t}{I_0}\right) \text{ and } V \propto I$$

Where: $A(c)$ is the absorbance of solution with concentration c, I_t is the intensity of light transmitted through the sample, I_0 is the intensity of light hitting the sample (from the orange LED) and V is the voltage measured across the red LED detector.

Using this, we can therefore rewrite the above equation in terms of voltages:

$$A(c) = -\log_{10}\left(\frac{V_c}{V_0}\right)$$

Where V_c is the voltage measured for solution of concentration c (equivalent to V_t) and V_0 is the voltage measured for the solvent, the control. In this experiment, we will use water.

Finally, we will define the ratio of the voltages in terms of transmittance, T:

$$T = \frac{V_c}{V_0}$$

Such that:

$$A(c) = -\log_{10}(T)$$

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Practical Work:

Health & Safety

Quinoline Yellow WS dye can stain clothes, surfaces and skin. Take care to avoid permanent staining of skin and clothes by wearing gloves and an apron. Wipe up spillages immediately.

Procedure: 50 Minutes

Materials:

- Gloves and Safety Specs
- 1x Red and 1x Blue LEDs
- Multimeter
- Lego® pieces
- 5 x 25 mL volumetric flasks and stoppers
- Plastic funnel
- 10 mL graduated pipette
- 50 mL graduated pipette
- Pipette filler
- 2 x red wires
- 1 x black wire
- 1 black and 1 red crocodile clip
- Red and black wires for the multimeter
- 100 Ω resistor
- Battery pack
- 7 x cuvettes
- Sample of 1 mM Quinoline Yellow WS dye
- Plastic pipettes
- 5 x 50 mL beaker
- 1 x 250 mL beaker
- Ruler and Graph Paper
- Permanent Marker
- Deionised Water
- Sample of Yellow Cough Sweets
- Stirrer Hot Plate and Stirrer Bar
- Mass Balance

Serial Dilution

First you will prepare solutions of known concentrations in order to calibrate your spectrophotometer. You have been supplied with stock solution of concentration 100 mM, which is the most concentrated solution under investigation. From this, you can prepare 6 solutions of varying concentrations by serial dilution.

1.0 mM	0.5 mM	0.25 mM	0.13 mM	0.06 mM	0.03 mM	0.0 mM

a) These values have been recorded to 2 d.p. and are not the exact values you can work out with a calculator. Why might this be?

Tip: Make sure you clean and rinse the pipette when moving between solutions of different concentrations. To do this, draw a little bit of deionised water into the pipette, and flush this into the cup you have designated for waste. Then, draw up a little bit of the solution you want to use, and flush this into the waste again. You can now use the pipette as normal. Pour some deionised water into a clean cup, and use this as your washing water.

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1. First, you will make the 0.5 mM Tartrazine solution.
2. Using a 10 mL graduated pipette and pipette filler, transfer 12.5 mL of the 1 mM dye solution to a 25 mL volumetric flask. You can do this is one go by filling the pipette up to the line marked -2.5 mL (at the top, above the 0 marker).
3. You may need to use the funnel, as the pipette may be too large to fit into the flask.
4. Rinse the funnel into the volumetric flask with a small amount of deionised water to ensure all the solution has been transferred.
5. Making sure you **do not go over the line**, top up with deionised water until the bottom of the meniscus touches the marked line. If you go over, you will need to start again.
6. To ensure the solution is homogenous, stopper the flask and invert it 5 times. This is the 0.5 mM solution.
7. Label the volumetric flask with the concentration of solution. This is important as some dilutions are not easily distinguishable with strong Quinoline Yellow WS dye!
8. Pour most of the 0.5 mM solution into a clean, labelled, 50 mL beaker.
9. Then, take 12.5 mL of this 0.5 mM solution using the **cleaned and rinsed** graduated pipette (see tip above), and transfer to a new 25 mL volumetric flask using a funnel if needed.
10. This is the 0.25 mM solution.
11. Continue with steps 2-9 for the remaining 3 concentrations, until you have made 5 new concentrations. Remember to remove 12.5 mL from the most dilute solution (lowest concentration) each time to make a new concentration.
12. Add deionised water to a 250 mL beaker to pipette from later on. This is the 0.0 µM solution.

b) Why do you need to rinse the pipette with the solution before using it?

c) Why do you need to put deionised water and stock solution into a separate container, rather than pipetting straight from the bottle?

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Calibration Plot

Next, you need to measure the absorbance of each solution. This is done by measuring the voltage across the red LED when each solution is placed in the spectrophotometer and the blue LED is on.

First, you need to measure the deionised water as a reference:

1. Add approximately 3 mL of deionised water into a cuvette using a plastic bulb pipette. The cuvette should be about 2/3 rds full.
2. If there is any liquid on the outside of the cuvette, make sure this is wiped off.
3. Place the cuvette in the spectrophotometer, making sure the blue/red LEDs are pointing through the **clear side with the arrow marked on it**, as the other two sides are slightly cloudy.
4. Turn on the blue LED by switching the battery pack to 'on'.
5. Turn on the multimeter, and make sure the central dial is pointing to the left-hand side '200m' (200 mV). If the multimeter is overloaded, change the setting to 2 V.
6. Allow the voltage to settle.
7. Note down the voltage, and fill out the table below.
8. Repeat for the other 6 solutions, using a **clean** cuvette each time, and fill out the table with V(c). Make sure you change the plastic pipette in-between solutions.
9. Make sure you turn off the LED when not in use.

Table 1: Concentration Calibration

Concentration of solution, $\mathbf{c}$ / mM	$\mathbf{V(c)}$ / [200m]V	$\mathbf{T = \frac{V(c)}{V(H_2O)}}$	$\mathbf{A(c) = -\log_{10}(T)}$
0.0 (water reference)			
0.031			
0.063			
0.13			
0.25			
0.50			
1.00			

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Determination of Unknown Concentration

Now you can measure the absorbance of the yellow cough sweets, which will be used later to determine its concentration.

c) Can you estimate the concentration of dye in the sweets by looking at the colour?

First, prepare your cough sweet solution. You may wish to begin this preparation before measuring the absorbances of the serial dilutions of quinoline yellow as the sweets may take longer to dissolve.

1. Remove three cough sweets from the packaging and record their combined mass.
2. Place the sweets in a 50 mL beaker and add 10 mL of deionised water and a stirrer bar.
3. With warming the mixture on a hotplate to 50 degrees Celsius, the sweets should take about ten minutes to dissolve. Ensure complete dissolution for later measurement!
4. Once the sweets have dissolved, carefully transfer the mixture to a 25 mL volumetric flask and dilute to the line. Rinse the contents of the beaker and funnel into the flask to ensure all product is in the volumetric flask.

Now, you need to clean your cuvettes.

1. Pour the solution in the cuvette into the waste beaker.
2. Add a few mL of deionised water, washing down the sides of the cuvette, and pour this into the waste beaker.
3. Then add a small amount of the cough sweet solution you will be testing into the cuvette, also washing down the sides of the cuvette. Then pour this into the waste as well

The cuvettes will now be ready to use for more testing.

1. Clean one of the plastic bulb pipettes
2. Pipette approximately 3 mL of the cough sweet solution into a clean cuvette with a **cleaned** bulb pipette. It should be about 2/3 rds full.
3. Place this into the spectrophotometer, making sure the outside is dry, and the clear sides (with the arrow on) are facing the LEDs.
4. Measure the voltage across the red LED as above, and note this down in the table on the previous page.

Table 2: Measurement of Unknown Concentration of Sports Drink

Concentration of solution, $\mathbf{c}$ / μM	$\mathbf{V(c)}$ / [200m]V	$\mathbf{T = \frac{V(c)}{V(H_2O)}}$	$\mathbf{A(c) = -\log_{10}(T)}$
Cough Sweets			

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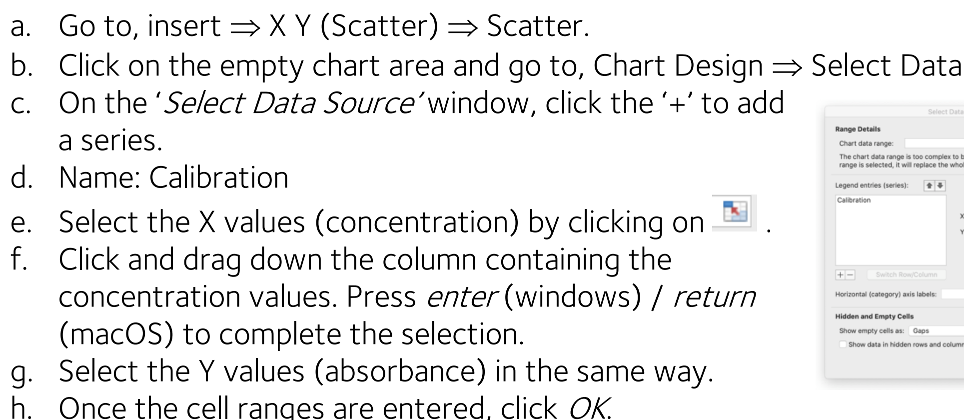
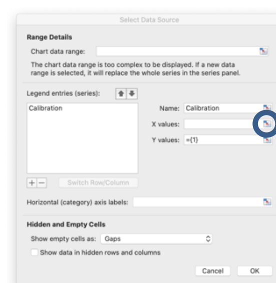
Post Lab Questions:

This part is much easier to complete using graphing software, but can also be done by hand with graph paper.

1. According to Beer's law, plotting $A(c)$ (sample absorbance) against c (concentration) makes a linear plot to which we can draw a line of best fit. The easiest way to do this is in Microsoft Excel. Do **not** include your value for the sports drink in the calibration plot.



   a. Go to, insert ⇒ X Y (Scatter) ⇒ Scatter.
   b. Click on the empty chart area and go to, Chart Design ⇒ Select Data
   c. On the '*Select Data Source*' window, click the '+' to add a series.
   d. Name: Calibration
   e. Select the X values (concentration) by clicking on .
   f. Click and drag down the column containing the concentration values. Press *enter* (windows) / *return* (macOS) to complete the selection.
   g. Select the Y values (absorbance) in the same way.
   h. Once the cell ranges are entered, click *OK*.
   i. To add a line of best fit, click on the chart area and right click (windows) / hold control (^) + click (macOS). Click on '*Add Trendline...*'
   j. The trendline should be a straight line.



2. Are there any values that don't fit? What could you do to fix this?
3. At high concentrations, this line often is not straight, and it will flatten out. Why might this happen?
4. Using your absorbance value, and the calibration plot above, calculate the concentration of dye in the cough sweets provided. Give your answer in in mol dm^{-3} and to a suitable number of significant figures.
5. The molar mass of Quinoline Yellow WS is 477.38 g mol^{-1}. What is the concentration of the dye in g dm^{-3}?
6. The EU regulations state that the safe limit of Quinoline Yellow WS dye is 0.5 mg per kg of bodyweight per day. Can you work out how many cough sweets you could safely consume in one day (on the basis of dye concentration)?

> Tip: This is easier if you include units in your calculations! Convert g to mg and dm^3 to mL before doing any calculations.

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